

# Package ‘AlleleShift’

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**Type** Package

**Title** Predict and Visualize Population-Level Changes in Allele Frequencies in Response to Climate Change

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**Description** Methods (<[doi:10.7717/peerj.11534](https://doi.org/10.7717/peerj.11534)>) are provided of calibrating and predicting shifts in allele frequencies through redundancy analysis ('vegan::rda()') and generalized additive models ('mgcv::gam()'). Visualization functions for predicted changes in allele frequencies include 'shift.dot.ggplot()', 'shift.pie.ggplot()', 'shift.moon.ggplot()', 'shift.waffle.ggplot()' and 'shift.surf.ggplot()' that are made put data sets that are prepared by helper functions for each visualization method. Examples in the documentation show how to prepare animated climate change graphics through a time series with the 'gganimate' package. Function 'amova.rda()' shows how Analysis of Molecular Variance can be directly conducted with the results from redundancy analysis.

**License** GPL-3

**Imports** vegan (>= 2.6-4), BiodiversityR (>= 2.15-4), adegenet

**Depends** R (>= 4.1.0)

**Suggests** poppr, mgcv, dplyr, ggplot2, GGally, ggforce, ggrepel, ggsci, gggibbous, gganimate, patchwork

**NeedsCompilation** no

**Repository** CRAN

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## Contents

amova.rda . . . . .	2
count.model . . . . .	4
Poptri.genind . . . . .	8
population.shift . . . . .	9

shift.dot.ggplot . . . . .	11
shift.moon.ggplot . . . . .	13
shift.pie.ggplot . . . . .	14
shift.surf.ggplot . . . . .	17
shift.waffle.ggplot . . . . .	21
VIF.subset . . . . .	22

<b>Index</b>	<b>24</b>
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amova.rda	<i>Analysis of Molecular Variance (AMOVA) via Redundancy Analysis</i>
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## Description

The main objective of the function is to illustrate how information on 'mean squares' can be extracted from redundancy analysis results. For balanced data sets, the final 'Phi' statistics are expected to be the same as those obtained with [poppr.amova](#). The function can only handle 1-level (population) and 2-level (eg, region/population or population/subpopulation) hierarchies.

## Usage

```
amova.rda(x, x.data)
```

## Arguments

x	Result of redundancy analysis fitted via <a href="#">rda</a> .
x.data	Data used to fit the redundancy analysis via <a href="#">ensemble.VIF.dataframe</a> .

## Details

A similar analysis of the relationship between AMOVA and RDA is given by Kindt (2020), also exploring relationships with the Multivariate Analysis of Variance Using Distance Matrices methods that are available via [adonis2](#). For discussions on AMOVA and its relationship with the matrix of squared Euclidean distances between individuals, see Michalakis and Excoffier (1996), Peakall et al. (1995) or Meirmans and Liu (2018) (the last article also shows how AMOVA can be expanded beyond haploid and diploid organisms). These authors provide the coefficients (equations 3, 4a and 4b) that need to be used with unbalanced numbers of individuals in populations and/or groups, which could then be used to obtain the exact AMOVA statistics from the estimated Mean Squares (there is no practical point in doing this as AMOVA is available via [poppr.amova](#)).

I have also cross-checked the results with GenAIEx 6.5 (Peakall and Smouse 2012), using the export function of [genind2genalex](#) to cross-check that RDA obtains the correct Sums-of-Squares and Mean-Squares. Important to note here is that the values in GenAIEx are obtained from only the upper part of the distance matrix, hence these are exactly 50 percent of the Sums-of-Squares from RDA.

Note that for diploid organisms, the option here is not to include the hierarchical level within individuals.

**Value**

The function proceeds with Analysis of Molecular Variance from data generated by redundancy analysis (RDA).

**Author(s)**

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

**References**

- Kindt R. 2020. Analysis of Molecular Variance (AMOVA) with vegan and BiodiversityR, including a graphical method to identify potential migrants. <https://rpubs.com/Roeland-KINDT>
- Meirmans PG and Liu S. 2018. Analysis of Molecular Variance (AMOVA) for Autopolyploids. *Front. Ecol. Evol.*, 23 May 2018. doi:10.3389/fevo.2018.00066
- Michalakis Y and Excoffier L. 1996. A Generic Estimation of Population Subdivision Using Distances Between Alleles With Special Reference for Microsatellite Loci. *Genetics* 142: 1061-1064.
- Peakall R, Smouse PE and Huff DR. 1995. Evolutionary implications of allozyme and RAPD variation in diploid populations of dioecious buffalograss (*Buchloe dactyloides* (Nutt.) Engelm.). doi:10.1111/j.1365294X.1995.tb00203.x.
- Peakall R and Smouse PE. 2012. GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research an update. doi:10.1093/bioinformatics/bts460.

**Examples**

```
## Not run:

library(poppr)
library(BiodiversityR) # also loads vegan

# Example with 1 level
data(nancycats)
names(alleles(nancycats)) <- locNames(nancycats) # small bug in this data set
nancycats2 <- missingno(nancycats, type = "loci", cutoff=0)
nancy.dist <- vegdist(nancycats2@tab, method="euc")

# Same method as in GenAlEx via the squared Euclidean distance
nancy.dist.amova <- poppr.amova(nancycats2, ~Population, within=FALSE,
                               dist=nancy.dist, squared=FALSE)
nancy.dist.amova

# Via vegan::rda
library(vegan)
nancycats.rda.data <- data.frame(Population=nancycats2@pop)

nancycats.rda.result <- rda(nancycats2@tab ~ Population, data=nancycats.rda.data)
nancycats.rda.result
amova.rda(nancycats.rda.result, nancycats.rda.data)

# Example with 2 levels
```

```

# Same example as for poppr::poppr.amova
data(Aeut)
strata(Aeut) <- other(Aeut)$population_hierarchy[-1]
agc <- as.genclone(Aeut)
agc
amova.result <- poppr.amova(agc, ~Pop/Subpop, within=FALSE)
amova.result

agc.rda.data <- data.frame(as.matrix(agc$other$population_hierarchy))
agc.rda.data[, 1] <- as.factor(agc.rda.data[, 1])
agc.rda.data[, 2] <- as.factor(agc.rda.data[, 2])
agc.rda.data[, 3] <- as.factor(agc.rda.data[, 3])
str(agc.rda.data)

agc.rda.result <- rda(agc@tab ~ Pop + Pop_Subpop, data=agc.rda.data)
agc.rda.result
amova.rda(agc.rda.result, agc.rda.data)

# Example with a balanced data set
# library(BiodiversityR)
data(warcom)
data(warenv)

warburgia.genind <- genind(warcom)
warburgia.genind@strata <- warenv
poppr.amova(warburgia.genind, ~ population, within=FALSE)

warburgia.rda <- rda(warcom ~ population, data=warenv)
warburgia.rda
amova.rda(warburgia.rda, warenv)

## End(Not run)

```

---

count.model

*Calibration and Prediction Functions for Changes in Allele Frequencies*


---

### Description

The functions implement a two-step calibration and prediction process for allele frequencies, whereby the second calibration step uses the predictions of the first step.

### Usage

```

count.model(genpop.data, env.data, permutations = 99,
            ordistep = FALSE, cca.model = FALSE)

count.pred(count.modeled, env.data)

```

```

freq.model(count.predicted)

freq.pred(freq.modeled, count.predicted)

freq.ggplot(freq.predicted, plot.best = TRUE, threshold = 0.50,
colour.Pop = TRUE, manual.colour.values = NULL,
xlim = c(0.0, 1.0), ylim = c(0.0, 1.0))

```

## Arguments

genpop.data	Data in the format of <a href="#">genpop</a> .
env.data	Descriptors of (bio-)climatic conditions for the populations, either for the baseline climate (to check calibrations) or future/past climates. It is expected that these are in the same sequence as the populations in the <code>genpop.data</code> .
permutations	Number of permutations as in <a href="#">anova.cca</a> .
ordistep	Check the results of <a href="#">ordistep</a> .
cca.model	Fit a CCA model with the minor allele frequency as in Blumstein et al. 2020.
count.modeled	Model fitted by <code>count.model</code> .
count.predicted	Predictions made by <code>count.pred</code> , expected to be predicted for the baseline climate as a second step in the calibration process.
freq.modeled	Model fitted by <code>freq.model</code> .
freq.predicted	Allele frequencies predicted by <code>freq.pred</code> .
plot.best	Plot the models with highest R2 (TRUE) or lowest R2 values (FALSE).
threshold	Threshold used to distinguish between the best and worst model.
colour.Pop	Colour populations differently (TRUE), or alleles differently (FALSE)
manual.colour.values	Manual specifications for colour values
xlim	limits of the x-axis
ylim	limits of the y-axis

## Details

These functions allow for an almost completely alternative workflow of predicting shifts in allele frequencies under climate change than the protocol developed by Blumstein et al. 2020. The methodology available here calibrates and predicts changes in allele frequencies via redundancy analysis (Blumstein et al. use canonical correspondence analysis) that were calibrated from allele counts (Blumstein et al. use allele frequencies) for all alleles (Blumstein et al. only calibrate the minor alleles) in a two-step calibration process (the second step via `freq.model` and `freq.predict` to ensure that predicted alleles are in the range of 0-1 is not included in protocol developed by Blumstein et al.). Other key differences in the methodology are that explanatory variables are expected to be bioclimatic variables (and not principal components as in Blumstein et al.) and that the input data is expected to be in the [genpop](#) format. Although a method to reduce the number of explanatory variables via [ordistep](#) is shown, I advise against reducing the explanatory variables

as this likely will reduce the explanatory power of the models, whereas explanatory power is the major objective of calibrating these functions. Motivations for the differences in methodologies of Blumstein et al. and the ones available in AlleleShift are explained by Kindt 2021.

Confidence intervals are calculated via `qt` with  $p=0.95$  and  $df=np$  (number of populations), although the GAM is fitted only once for all the alleles and all the populations (that was my choice to reduce overfitting the baseline data).

The 'darkolivegreen4'-coloured reference lines shown via `freq.ggplot` correspond to a 1:1 relationship (full line) and 1:0.95 and 1:1.05 relationships (dashed lines).

## Value

The functions enable calibration and prediction of allele frequencies.

## Author(s)

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

## References

Blumstein et al. 2020. Protocol for Projecting Allele Frequency Change under Future Climate Change at Adaptive-Associated Loci. [doi:10.1016/j.xpro.2020.100061](https://doi.org/10.1016/j.xpro.2020.100061)

Kindt R. 2021. AlleleShift: An R package to predict and visualize population-level changes in allele frequencies in response to climate change. *PeerJ* 9:e11534. [doi:10.7717/peerj.11534](https://doi.org/10.7717/peerj.11534)

Brauer CJ., Sandoval-Castillo J, Gates K et al. 2023, Natural hybridization reduces vulnerability to climate change. *Nat. Clim. Chang.* 13: 282-289, [doi:10.1038/s41558022015851](https://doi.org/10.1038/s41558022015851)

## See Also

[VIF.subset](#), [population.shift](#), [amova.rda](#)

## Examples

```
# A typical work flow would consist of:

# 1. Reduce the number of explanatory variables

data(Poptri.baseline.env)
data(Poptri.future.env)

VIF.select <- VIF.subset(Poptri.baseline.env,
                        keep=c("MAT", "CMI"),
                        cor.plot=TRUE)

VIF.select$VIF$vars.included

baseline.env <- Poptri.baseline.env[, VIF.select$VIF$vars.included]
summary(baseline.env)
```

```
future.env <- Poptri.future.env[, VIF.select$VIF$vars.included]

# 2. Create the genpop object

data(Poptri.genind)
Poptri.genpop <- adegenet::genind2genpop(Poptri.genind)

# Get to know the populations and the alleles
poppr::poppr(Poptri.genind)
adegenet::makefreq(Poptri.genpop)

# 3. Calibrate the models

# Note that the ordistep procedure is not needed
# CCA model only to compare results with those by Blumstein et al. 2020
Poptri.count.model <- count.model(Poptri.genpop,
                                  env.data=baseline.env,
                                  ordistep=TRUE,
                                  cca.model=TRUE)

Poptri.pred.baseline <- count.pred(Poptri.count.model, env.data=baseline.env)
head(Poptri.pred.baseline)

Poptri.freq.model <- freq.model(Poptri.pred.baseline)
Poptri.freq.baseline <- freq.pred(Poptri.freq.model,
                                  count.predicted=Poptri.pred.baseline)

head(Poptri.freq.baseline)

# 4. Check how well the models predict baseline allele frequencies

# Populations are split in those with R2 > 0.50 and those with R2 < 0.50
# Better populations
plotA1 <- freq.ggplot(Poptri.freq.baseline,
                     plot.best=TRUE,
                     ylim=c(0.0, 0.8))
plotA1

# Populations with low R2
manual.colour.values1 <- ggsci::pal_npg()(5)

plotB1 <- freq.ggplot(Poptri.freq.baseline,
                     plot.best=FALSE,
                     manual.colour.values=manual.colour.values1,
                     xlim=c(0, 0.5),
                     ylim=c(0, 0.25))
plotB1

# Colouring by alleles

plotA2 <- freq.ggplot(Poptri.freq.baseline,
                     colour.Pop=FALSE,
```

```

        plot.best=TRUE,
        ylim=c(0.0, 0.8),
        manual.colour.values=manual.colour.values1)

plotA2

plotB2 <- freq.ggplot(Poptri.freq.baseline,
                    colour.Pop=FALSE,
                    plot.best=FALSE,
                    manual.colour.values=manual.colour.values1,
                    xlim=c(0, 0.5),
                    ylim=c(0, 0.25))

plotB2

# 5. Predict future allele frequencies

Poptri.pred.future <- count.pred(Poptri.count.model, env.data=future.env)
head(Poptri.pred.future)

Poptri.freq.future <- freq.pred(Poptri.freq.model,
                               count.predicted=Poptri.pred.future)

# The key results are variables 'Allele.freq' representing the baseline allele frequencies
# and variables 'Freq.e2', the predicted frequency for the future/ past climate.
# Variable 'Freq.e1' is the predicted allele frequency in step 1

head(Poptri.freq.future)

# 5. Visualize the changes
# See functions shift.dot.ggplot, shift.pie.ggplot, shift.moon.ggplot,
# shift.waffle.ggplot and shift.surf.ggplot

```

---

Poptri.genind

*Example data sets for Populus trichocarpa.*

---

## Description

These data sets include the same data that were used by Blumstein et al. 2020 to document their protocol. Genomic data has been processed further into the [genind](#) format. Data sets Poptri.freq.baseline, Poptri.freq.future and Poptri.1985to2085 were obtained with the calibration and prediction methods of this package. They are included to make the documentation of the various plotting methods shorter, but also show the types of results someone can obtain.

## Usage

```

data(Poptri.genind)
data(Poptri.baseline.env)

```



```

data(Poptri.future.env)
data(Poptri.loc)
data(Poptri.freq.baseline)
data(Poptri.freq.future)
data(Poptri.1985to2085)

```

## References

Blumstein et al. 2020. Protocol for Projecting Allele Frequency Change under Future Climate Change at Adaptive-Associated Loci. [doi:10.1016/j.xpro.2020.100061](https://doi.org/10.1016/j.xpro.2020.100061)

## Examples

```
data(Poptri.genind)
```

---

population.shift	<i>Shifts of Populations in Environmental Space as Response to Climate Change</i>
------------------	---

---

## Description

The function plots the locations of each population in baseline and future climates. Arrows indicate the shifts in positions of the populations.

## Usage

```

population.shift(baseline.env.data, future.env.data,
option=c("PCA", "RDA"), vector.multiply=1)

environmental.novel(baseline.env.data, future.env.data)

```

## Arguments

baseline.env.data	Baseline (bio-)climatic conditions for the populations.
future.env.data	Changed (bio-)climatic conditions in future/past for the populations.
option	Should an explanatory variable corresponding to the climate period be used by <a href="#">rda</a> .
vector.multiply	Multiplier for vector scores in the ordination diagrams.

**Details**

See Kindt (2020) for alternative methods of generating ordination diagrams via `vegan`, `BiodiversityR` and `ggplot2`.

Function `environmental.novel` identifies populations with future (or past) environmental conditions that are outside the baseline range. The function further calculates the probability of observing the future condition via `pnorm` with the mean and standard deviation from the baseline conditions. Where one or several variables are outside the baseline range, data are provided for the variable with the smallest probability.

**Value**

The main function generates an ordination diagram that depicts changes between baseline and future/past conditions for the populations.

**Author(s)**

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

**References**

Kindt R. 2020. Ordination graphs with `vegan`, `BiodiversityR` and `ggplot2`. <https://rpubs.com/Roeland-KINDT>

**Examples**

```
data(Poptri.baseline.env)
data(Poptri.future.env)

environmental.novel(Poptri.baseline.env, Poptri.future.env)
# as if for past climates
environmental.novel(Poptri.future.env, Poptri.baseline.env)

VIF.select <- VIF.subset(Poptri.baseline.env,
                        keep=c("MAT", "CMI"),
                        cor.plot=FALSE)

VIF.select$vars.included

baseline.env <- Poptri.baseline.env[, VIF.select$vars.included]
future.env <- Poptri.future.env[, VIF.select$vars.included]

environmental.novel(baseline.env, future.env)

plotA <- population.shift(baseline.env,
                          future.env,
                          option="PCA")

plotA

plotB <- population.shift(baseline.env,
```

```
future.env,  
option="RDA")  
plotB
```

---

shift.dot.ggplot      *Shifts of Allele Frequencies as Response to Climate Change*

---

### Description

The function shows changes in allele frequencies between the baseline and future/past climate.

### Usage

```
shift.dot.ggplot(freq.future,  
mean.change = FALSE, change.FUN = stats::median,  
baseline.colour = "black",  
future.colour = "dodgerblue3",  
manual.colour.values=c("firebrick3", "chartreuse4"))
```

### Arguments

freq.future	Result from AlleleShift::predFreq..
mean.change	Aggregate changes among alleles.
change.FUN	Function used the aggregate changes.
baseline.colour, future.colour, manual.colour.values	Colours to be used in the plots.

### Value

The function generates a ggplot that depicts changes between baseline and future/past allele frequencies of the populations.

### Author(s)

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

### See Also

[shift.pie.ggplot](#), [shift.moon.ggplot](#), [shift.waffle.ggplot](#), [shift.surf.ggplot](#)

**Examples**

```

# The data can be obtained via the count.model and freq.model calibrations.
# These procedures are not repeated here.
data(Poptri.freq.future)

ggdot1 <- shift.dot.ggplot(Poptri.freq.future)
ggdot1

# create an animation

## Not run:

library(ggplot2)
library(gganimate)
library(gifski)

# The data is an interpolation and extrapolation between the baseline and future climate.
# For actual application, interpolate between climate data from available sources

data(Poptri.1985to2085)

ggdot.all <- ggplot(data=Poptri.1985to2085, group=Decade) +
  scale_y_continuous(limits=c(-0.1, 1.1),
                    breaks=c(0.0, 0.25, 0.5, 0.75, 1.0)) +
  geom_errorbar(aes(x=Pop, ymin=LCL, ymax=UCL),
               colour="grey30", width=0.8, show.legend=FALSE) +
  geom_segment(aes(x=Pop, y=Allele.freq, xend=Pop, yend=Freq.e2, colour=increasing),
              size=1.2) +
  geom_point(aes(x=Pop, y=Allele.freq),
             colour="black", size=10, alpha=0.7) +
  geom_point(aes(x=Pop, y=Freq.e2),
            colour="dodgerblue3", size=10, alpha=0.7) +
  coord_flip() +
  xlab(element_blank()) +
  ylab("Allele frequencies") +
  theme(panel.grid.minor = element_blank()) +
  labs(colour="Future change in allele frequencies") +
  scale_colour_manual(values=c("firebrick3", "chartreuse4"),
                     labels=c("decreasing", "increasing")) +
  theme(axis.text.x=element_text(angle=90, vjust=0.5, size=10)) +
  theme(legend.position="top") +
  facet_grid(~ Allele, scales="free")

ggdot.all

ggdot.anim <- ggdot.all +
  transition_states(as.factor(Decade), transition_length = 10, state_length = 100) +
  labs(title = "Decade: {closest_state}s")

ggdot.anim2 <- animate(ggdot.anim, fps=5, width=1280, height=720)

getwd()

```

```
anim_save(filename="Allele shift animation.gif", animation=ggdot.anim2)
```

```
## End(Not run)
```

---

```
shift.moon.ggplot      Shifts of Allele Frequencies as Response to Climate Change
```

---

## Description

The function shows changes in allele frequencies between the baseline and future/past climate.

## Usage

```
shift.moon.ggplot(baseline.moon, future.moon,
  manual.colour.values = c("white", "grey", "firebrick3", "chartreuse4"),
  manual.colour.codes = c("A baseline ", "B", "A decreasing", "A increasing"))
```

```
moon.waxer(freq.in, sort.index= "Pop.index",
  mean.change = FALSE, change.FUN = stats::median,
  freq.focus = "Allele.freq",
  ypos = 0, right = TRUE)
```

## Arguments

baseline.moon, future.moon	Result from AlleleShift::moon.waxer.
manual.colour.values	Colours to be used in the plot.
manual.colour.codes	Sequence for the manual colour values.
freq.in	Result from AlleleShift::predFreq.
sort.index	Sequence of the populations in the plot.
mean.change	Aggregate changes among alleles.
change.FUN	Function used the aggregate changes.
freq.focus	Allele frequency for which to calculate statistics, either 'Allele.freq' or 'Freq.e2'.
ypos, right	Arguments used for plotting, mainly as in <a href="#">geom_moon</a>

## Value

The function generates a ggplot that depicts changes between baseline and future/past allele frequencies of the populations.

**Author(s)**

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

**See Also**

[shift.dot.ggplot](#), [shift.pie.ggplot](#), [shift.waffle.ggplot](#), [shift.surf.ggplot](#)

**Examples**

```
# The data can be obtained via the count.model and freq.model calibrations.
# These procedures are not repeated here.
data(Poptri.freq.baseline)
data(Poptri.freq.future)

Poptri.baseline.moon <- moon.waxer(Poptri.freq.baseline,
                                  sort.index="Latitude.index")

Poptri.future.moon <- moon.waxer(Poptri.freq.future,
                                 sort.index="Latitude.index",
                                 freq.focus="Freq.e2",
                                 ypos=1)

ggmoon1 <- shift.moon.ggplot(Poptri.baseline.moon,
                             Poptri.future.moon)

ggmoon1
```

---

shift.pie.ggplot

*Shifts of Allele Frequencies as Response to Climate Change*

---

**Description**

The function shows changes in allele frequencies between the baseline and future/past climate.

**Usage**

```
shift.pie.ggplot(baseline.pie, future.pie,
  manual.colour.values = c("black", "grey", "firebrick3", "chartreuse4"),
  manual.colour.codes = c("A baseline ", "B", "A decreasing", "A increasing"))

pie.baker(freq.in, sort.index= "Pop.index",
  mean.change = FALSE, change.FUN = stats::median,
  freq.focus = "Allele.freq", ypos = 0,
  r0 = 0.1, r = 0.5, focus = 0.2 )
```

**Arguments**

baseline.pie, future.pie	Result from AlleleShift::pie.baker.
manual.colour.values	Colours to be used in the plot.
manual.colour.codes	Sequence for the manual colour values.
freq.in	Result from AlleleShift::predFreq.
sort.index	Sequence of the populations in the plot.
mean.change	Aggregate changes among alleles.
change.FUN	Function used the aggregate changes.
freq.focus	Allele frequency for which to calculate statistics, either 'Allele.freq' or 'Freq.e2'.
ypos, r0, r, focus	Arguments used for plotting, mainly as in <a href="#">geom_arc_bar</a>

**Value**

The function generates a ggplot that depicts changes between baseline and future/past allele frequencies of the populations.

**Author(s)**

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

**See Also**

[shift.dot.ggplot](#), [shift.moon.ggplot](#), [shift.waffle.ggplot](#), [shift.surf.ggplot](#)

**Examples**

```
# The data can be obtained via the count.model and freq.model calibrations.
# These procedures are not repeated here.
data(Poptri.freq.baseline)
data(Poptri.freq.future)

Poptri.baseline.pie <- pie.baker(Poptri.freq.baseline, r0=0.1,
                               sort.index="Latitude.index")

Poptri.future.pie <- pie.baker(Poptri.freq.future, r0=0.1,
                              freq.focus="Freq.e2",
                              sort.index="Latitude.index",
                              ypos=1)

ggpie1 <- shift.pie.ggplot(Poptri.baseline.pie,
                          Poptri.future.pie)
ggpie1
```

```

# create an animation

## Not run:

library(ggplot2)
library(ggforce)
library(gganimate)
library(gifski)
library(transformr)

# The data is an interpolation and extrapolation between the baseline and future climate.
# For actual application, interpolate between climate data from available sources

data(Poptri.1985to2085)

decades <- sort(unique(Poptri.1985to2085$Decade))

for (d in 1:length(decades)) {
  decade.focal <- decades[d]
  decade.data <- Poptri.1985to2085[Poptri.1985to2085$Decade == decade.focal, ]
  decade.pie <- pie.baker(decade.data, r0=0.1,
                          freq.focus="Freq.e2",
                          sort.index="Latitude.index",
                          ypos=1)
  decade.pie <- cbind(Decade=rep(decade.focal, nrow(decade.pie)), decade.pie)

  if (d == 1) {
    future.pies <- decade.pie
  }else{
    future.pies <- rbind(future.pies, decade.pie)
  }
}

np <- length(unique(Poptri.baseline.pie$Pop))
manual.colour.values <- c("black", "grey", "firebrick3", "chartreuse4")

ggpie.all <- ggplot(data=future.pies, group=Decade) +
  scale_x_continuous(limits=c(0.5, np+0.5),
                    breaks=seq(from=1, to=np, by=1),
                    labels=levels(Poptri.baseline.pie$Pop)) +
  geom_arc_bar(data=Poptri.baseline.pie,
              aes(x0=sort.index, y0=ypos, r0=r0, r=0.4,
                 start=start, end=end, fill=colour),
              size=0.04, alpha=1, colour="snow1") +
  geom_arc_bar(data=future.pies,
              aes(x0=sort.index, y0=ypos, r0=r0, r=0.4,
                 start=start, end=end, fill=change.colour),
              size=0.04, alpha=1, colour="snow1") +
  geom_point(data=subset(future.pies, increasing==TRUE),
            aes(x=sort.index, y=ypos),
            size=5, shape=21, fill=manual.colour.values[4], stroke=0.03, show.legend=FALSE) +
  geom_point(data=subset(future.pies, increasing==FALSE),
            aes(x=sort.index, y=ypos),

```



```

        size=5, shape=21, fill>manual.colour.values[3], stroke=0.03, show.legend=FALSE) +
coord_flip() +
xlab(element_blank()) +
ylab(element_blank()) +
labs(fill=" ") +
scale_fill_manual(values>manual.colour.values,
                  labels=c("A baseline ", "B", "A decreasing", "A increasing")) +
theme(panel.grid = element_blank()) +
theme(axis.text.x=element_blank()) +
theme(axis.ticks.x = element_blank()) +
theme(legend.position="top") +
facet_grid( ~ Allele, scales="free")

ggpie.all

# note this will take quite a while!
ggpie.anim <- ggpie.all +
  transition_states(as.factor(Decade), transition_length = 10, state_length = 100) +
  labs(title = "Decade: {closest_state}s")

ggpie.anim2 <- animate(ggpie.anim, fps=5, width=1280, height=720)

getwd()
anim_save(filename="Allele shift pie animation.gif", animation=ggpie.anim2)

## End(Not run)

```

---

 shift.surf.ggplot

*Shifts of Allele Frequencies as Response to Climate Change*


---

## Description

The function shows changes in allele frequencies between the baseline and future/past climate.

## Usage

```

shift.surf.ggplot(freq.future,
Allele.focus=unique(freq.future$Allele)[1],
freq.focus="Allele.freq",
xcoord="LON", ycoord="LAT",
mean.change = FALSE, change.FUN = stats::median,
manual.colour.values = c("firebrick3", "chartreuse4"),
...)

```

**Arguments**

freq.future	Result from AlleleShift::predFreq.
freq.focus	Selection of the Allele.
Allele.focus	Selection of the frequency.
xcoord, ycoord	Geographical coordinates of the populations.
mean.change	Aggregate changes among alleles.
change.FUN	Function used the aggregate changes.
manual.colour.values	Colours to be used in the plot.
...	Options for <code>ordisurf</code> .

**Details**

Populations are plotted in geographical space via ordination plotting methods, which is suitable as fixed coordinate systems are recommended both in ordination diagrams and maps. See Kindt (2020) for alternative methods of generating ordination diagrams via `vegan`, `BiodiversityR` and `ggplot2`.

Kindt (2021) shows how a STAMEN baseline map can be used to produce high resolution images via the `ggmap` package.

**Value**

The function generates a `ggplot` that depicts changes between baseline and future/past allele frequencies of the populations.

**Author(s)**

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

**References**

Kindt R. 2020. Ordination graphs with `vegan`, `BiodiversityR` and `ggplot2`. <https://rpubs.com/Roeland-KINDT>

Kindt, R. 2021. Plotting smoothed surface diagrams of allele frequencies obtained from `AlleleShift` on a baseline map via `ggmap`. <https://rpubs.com/Roeland-KINDT>

**See Also**

[shift.dot.ggplot](#), [shift.pie.ggplot](#), [shift.moon.ggplot](#), [shift.waffle.ggplot](#)

**Examples**

```
# The data can be obtained via the count.model and freq.model calibrations.
# These procedures are not repeated here.
data(Poptri.freq.baseline)
data(Poptri.freq.future)

# Plots for the first allele
```

```

# Symbols and colours indicate future change (green, ^ = future increase)
# Symbol size reflects the frequency in the climate shown

# Baseline climate
plotA <- shift.surf.ggplot(Poptri.freq.future,
                          xcoord="Long", ycoord="Lat",
                          Allele.focus=unique(Poptri.freq.future$Allele)[1],
                          freq.focus="Allele.freq")
plotA

# Future/past climate
plotB <- shift.surf.ggplot(Poptri.freq.future,
                          xcoord="Long", ycoord="Lat",
                          Allele.focus=unique(Poptri.freq.future$Allele)[1],
                          freq.focus="Freq.e2")
plotB

# Plots for the fifth allele

# Baseline climate
plotC <- shift.surf.ggplot(Poptri.freq.future,
                          xcoord="Long", ycoord="Lat",
                          Allele.focus=unique(Poptri.freq.future$Allele)[5],
                          freq.focus="Allele.freq")
plotC

# Future climate
plotD <- shift.surf.ggplot(Poptri.freq.future,
                          xcoord="Long", ycoord="Lat",
                          Allele.focus=unique(Poptri.freq.future$Allele)[5],
                          freq.focus="Freq.e2")
plotD

# create an animation

## Not run:

library(ggplot2)
library(ggforce)
library(gganimate)
library(gifski)
library(transformr)

# The data is an interpolation and extrapolation between the baseline and future climate.
# For actual application, interpolate between climate data from available sources

data(Poptri.1985to2085)

Poptri.1985to2085$xcoord <- Poptri.1985to2085$Long
Poptri.1985to2085$ycoord <- Poptri.1985to2085$Lat

alleles <- sort(unique(as.character(Poptri.1985to2085$Allele)))
future.data <- Poptri.1985to2085[Poptri.1985to2085$Allele == alleles[1], ]

```

```

decades <- sort(unique(future.data$Decade))

for (d in 1:length(decades)) {
  decade.focal <- decades[d]
  decade.data <- future.data[future.data$Decade == decade.focal, ]
  plotLONLAT <- vegan::ordiplot(decade.data[, c("xcoord", "ycoord")])
  surfAllele <- BiodiversityR::ordisurfgrid.long(vegan::ordisurf(plotLONLAT, y=decade.data$Freq.e2))
  decade.surf <- cbind(Decade=rep(decade.focal, nrow(surfAllele)), surfAllele)
  if (d == 1) {
    future.surfs <- decade.surf
  }else{
    future.surfs <- rbind(future.surfs, decade.surf)
  }
}

# The function above will not be able to predict far into the future.
# The results obtained (future.surfs) can still be used for plotting.

ggsurf.all <- ggplot(data=future.surfs, group=Decade) +
  geom_contour_filled(aes(x=x, y=y, z=z),
    breaks=seq(from=0.0, to=1.05, by=0.05)) +
  geom_point(data=subset(future.data, Decade==Decade),
    aes(x=xcoord, y=ycoord, size=Freq.e2, shape=increasing),
    colour="red", alpha=0.8, stroke=1.5, show.legend=FALSE) +
  xlab(element_blank()) +
  ylab(element_blank()) +
  labs(fill=alleles[1]) +
  scale_fill_viridis_d() +
  scale_colour_manual(values=c("firebrick3", "chartreuse4"),
    guide=FALSE) +
  scale_size_area(max_size=6) +
  scale_shape_manual(values=c(6, 2)) +
  theme(panel.grid = element_blank()) +
  theme(axis.text= element_blank()) +
  theme(axis.ticks = element_blank()) +
  theme(legend.title = element_text(size=9)) +
  theme(legend.text = element_text(size=8)) +
  coord_fixed()

ggsurf.all

ggsurf.anim <- ggsurf.all +
  transition_states(as.factor(Decade), transition_length = 10, state_length = 100) +
  labs(title = "Decade: {closest_state}s")

ggsurf.anim2 <- animate(ggsurf.anim, fps=5, width=1280, height=720)

getwd()
anim_save(filename="Allele shift surf animation.gif", animation=ggsurf.anim2)

## End(Not run)

```

---

shift.waffle.ggplot     *Shifts of Allele Frequencies as Response to Climate Change*

---

### Description

The function shows changes in allele frequencies between the baseline and future/past climate.

### Usage

```
shift.waffle.ggplot(future.waffle,  
manual.colour.values = c("black", "grey", "firebrick3", "chartreuse4"),  
manual.colour.codes = c("A baseline ", "B", "A decreasing", "A increasing"))  
  
waffle.baker(freq.in, sort.index = "Pop.index",  
mean.change = FALSE, change.FUN = stats::median)
```

### Arguments

future.waffle	Result from AlleleShift::waffle.baker.
manual.colour.values	Colours to be used in the plot.
manual.colour.codes	Sequence for the manual colour values.
freq.in	Result from AlleleShift::predFreq.
sort.index	Sequence of the populations in the plot.
mean.change	Aggregate changes among alleles.
change.FUN	Function used the aggregate changes.

### Details

Although a package ggwaffle exists, I opted to bake my own waffles (possibly the Belgian in me.). As a separate row is created for each square/rectangle of the waffle, the resulting data is quite large. Hence trying to animate this is probably a bad idea (unless you want to make some real waffles while your computer is busy).

### Value

The function generates a ggplot that depicts changes between baseline and future/past allele frequencies of the populations.

### Author(s)

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

**Examples**

```
# The data can be obtained via the count.model and freq.model calibrations.
# These procedures are not repeated here.
data(Poptri.freq.baseline)
data(Poptri.freq.future)

Poptri.future.waffle <- waffle.baker(Poptri.freq.future,
                                     sort.index="Latitude.index")

ggwaffle1 <- shift.waffle.ggplot(Poptri.future.waffle)
ggwaffle1
```

---

VIF.subset

*Reduce the Climate Data based on Variance Inflation Factor Analysis.*


---

**Description**

Through Variance Inflation Factor (VIF) analysis, a subset of variables is identified where all variables have VIF below a predefined threshold.

**Usage**

```
VIF.subset(data, VIF.max, keep=NULL, silent=FALSE, cor.plot=TRUE)
```

**Arguments**

data	(Bio)-Climatic or environmental descriptors of the populations
VIF.max	Maximum Variance Inflation Factor as in <a href="#">ensemble.VIF.dataframe</a> that is used internally.
keep	Variables to keep as in as in <a href="#">ensemble.VIF.dataframe</a> .
silent	Limit the verbose output as in <a href="#">ensemble.VIF.dataframe</a> .
cor.plot	Generate a correlation matrix for the final subset via <a href="#">ggcorr</a> .

**Value**

The function returns information on a subset of variables where VIF is below a pre-defined threshold.

**Author(s)**

Roeland Kindt (World Agroforestry, CIFOR-ICRAF)

**Examples**

```
data(Poptri.baseline.env)

# error as many variables are highly correlated
# VIF.subset(Poptri.baseline.env)

VIF.subset(Poptri.baseline.env, keep=c("MAT", "CMI"), cor.plot=TRUE)
```

# Index

## \* datasets

- Poptri.genind, 8
- adonis2, 2
- amova.rda, 2, 6
- anova.cca, 5
- count.model, 4
- count.pred (count.model), 4
- ensemble.VIF.dataframe, 2, 22
- environmental.novel (population.shift), 9
- freq.ggplot (count.model), 4
- freq.model (count.model), 4
- freq.pred (count.model), 4
- genind, 8
- genind2genalex, 2
- genpop, 5
- geom\_arc\_bar, 15
- geom\_moon, 13
- ggcorr, 22
- moon.waxer (shift.moon.ggplot), 13
- ordistep, 5
- ordisurf, 18
- pie.baker (shift.pie.ggplot), 14
- pnorm, 10
- poppr.amova, 2
- Poptri.1985to2085 (Poptri.genind), 8
- Poptri.baseline.env (Poptri.genind), 8
- Poptri.freq.baseline (Poptri.genind), 8
- Poptri.freq.future (Poptri.genind), 8
- Poptri.future.env (Poptri.genind), 8
- Poptri.genind, 8
- Poptri.loc (Poptri.genind), 8
- population.shift, 6, 9
- qt, 6
- rda, 2, 9
- shift.dot.ggplot, 11, 14, 15, 18
- shift.moon.ggplot, 11, 13, 15, 18
- shift.pie.ggplot, 11, 14, 14, 18
- shift.surf.ggplot, 11, 14, 15, 17
- shift.waffle.ggplot, 11, 14, 15, 18, 21
- VIF.subset, 6, 22
- waffle.baker (shift.waffle.ggplot), 21